CAP #2266301 CLIA #05D0542989

Molecular Pathology Genomics Core Dept. of Pathology & Lab Medicine Children's Hospital Los Angeles genomics@chla.usc.edu



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Next Generation and Sanger Sequencing Submission Form

Date of Submission:

Genomics Core Use Only-Project#: Sample Location: Received By: VMSR Sample ID:

Section 1: Contact Information

Principal Investigator:	Contact Person:
Department:	Phone #:
Institution:	Email:
Phone #:	
Email:	Human Subjects Approval:
Affiliation: If other, please specify:	IRB#:
Billing	
Billing Contact:	Account #:
Phone #:	
Email:	
PI Signature (Indicates financial commitment for services provi	ided):

Section 2: Service Information

Service Type:

For Target Region Sequencing, please specify:

Next-Generation Sequencing Method:

Multiplex □Yes □No

If yes, # of samples per chip:

Section 3: Sequencing and Fragment Sizing Sample Information

*For Sanger Sequencing, please only complete section 4.								
#	Sample Name	Sample Type	Concentration (ng/µl)	Volume (µI)	Well # on Gel Image			
1.	·							
2.								
3.								
4.								
5.								
6.								
7.								
8.								
9.								
10.								
11.								
12.								
13.								
14.								
15.								
16.								
17.								
18.								
19.								
20.								

*Please attach gel image

Section 4: Sanger Sequencing Sample Information												
Section A- REQUIRED				Section B or C - REQUIRED								
	Section A				Section B			OR	OR Section C			
	Variant Name	HUGO Gene	Nucleotide Reference	Nucleotide Variant	Zygosity (Het,Homo,Hemi)	Genome Build #	Chr #	Genomic Position - Start	Genomic Position - End		Transcript ID (NM#)	cDNA Position #
SNV example	MSH2_var1	MSH2	С	A	Heterozygous	36	6	572433	572433		NM_425	c.255
DEL example	JAG1_var1	JAG1	TG		Heterozygous	37	20	10633150	10633150		NM_000214.2	c.852_853
1												
2												
3												
4												
5												
6												
7												
8												
9												
10												

т	, MDO		Description	Fee Applied		
Туре			Description			
Initial Consultation (30 minutes) > Brie			ef discussion and overview of project scope.		NO; Initial 30 minutes	
LUTODAL			vide basic training with standard analysis tool ining suitable to next generation sequencing da	ata.	YES	
A La Carte > Opt			Ilyze the data for investigator. ion only available to researchers generating no eration sequencing data within MPG Core.	YES		
Collaborative			ended for large scale projects difficult to asses ure of analysis required.	S	NO; Requires percentage of effor within grant proposal	
Data Delivera	ble Format					
Alignment Genon	ne, (i.e. hg19):					
Variant Calling Analysis Standard Deliverables Includes: Ion Torrent .basecaller.bam .bai .vcf Includes: Ion Torrent .bam .bai .vcf Illumina .bai .bai .bai .bai .vcf Ison torrent .bai .vcf		erables	(Additional fees apply)		Custom Deliverable (Additional fees apply)	
		Includes: .fastq .sam .sai FastQC output BAM QC output Coverage report (Coverage may be standard output from Torrent Suite, CASAVA or our custom scripts) Genotyping QC report/filtered VCF Copy number analysis report	Any changes to the standard workflow are considered a custom analysis. Plea contact us for additional information.			
RNA Sequencing	Standard Delive	erables	 Extended Deliverables (Additional fees apply) 	(A	ustom Deliverable Additional fees apply)	
	Includes: Ion Torrent .basecaller.bam .bam .bai Illumina .bcl .fastq .bam .bai		Includes: .fastq .sam .sai FastQC output BAM QC output Coverage report	are co	nanges to the standard workflow onsidered a custom analysis. Please ct us for additional information.	

Additional Notes:

Laboratory:

Bioinformatics

Sample Requirements and Shipping Instructions

Sample Requirements

	Genomic DNA Requirements	Total RNA Requirements	PCR Amplicon	Data Only				
Sample Type Purified DNA (PureGene Extraction Preferred); 260/ ratio 1.75-2.00		Purified RNA; RNA Integrity Number(RIN) 8.0- 10.0	"Load Only" reactions should be submitted dry and ready to reconstitute for loading. All other samples must be amplified with PCR primers including M13 universal primer sequences at 5' ends. M13 fwd primer sequence: 5'-TGTAAAACGACGGCCAGT-3' M13 rev primer sequence: 5'-CAGGAAACAGCTATGACC-3'	Media/hard drive				
Collection Type	Screw cap tube	Screw cap tube	Screw cap tube	Encrypted. Retain original data source; lab not responsible for damage during shipping.				
Volume and/or suspended in suspende		2.0ug @ 100ng/ul suspended in nuclease free water.	Submit 25ul of your PCR product for sequencing or 10ul of your diluted PCR product for Fragment Analysis. For Fragment Analysis, amplicons must be between 35 and 500bp and labeled with FAM, VIC, NED, or PET; (the core suggests running a serial dilution for each locus if you are unfamiliar with the correct dilution of your amplicons).	N/A				
Labeling of Collection Type	Ensure label on tube is legible and matches information on submission sheet. Sample names should be alphanumeric with no more than 6 characters. Each tube must have a unique sample name.							
Shipping Instru	ctions							
Packaging Instructions If needed, sample packaging instructions are available as a pdf from FEDEX: Biological Substance: <u>http://images.fedex.com/us/packaging/guides/UN3373_fxcom.pdf</u> Media/Hard Drive: <u>http://images.fedex.com/us/packaging/guides/Computer_fxcom.pdf</u>								
Shipping	Ship at ambient temperature	Ship on dry ice	Ship on ice packs	Send on encrypted media/hard drive. Provide encryption method and key				
Instructions	Ship via overnight courier(FedEx,UPS)	Ship via overnight courier(FedEx,UPS)	Ship via overnight courier(FedEx,UPS)	via email. Ship via overnight courier(FedEx,UPS)				
Shipping Address	Molecular Pathology Genomics Core Children's Hospital Los Angeles 4650 Sunset Boulevard, MS #103 Los Angeles, California 90027							